CHROMATOGRAPHY

emp BIOTECH

CentriPure MINI Spin Columns

Unique conical column for greater separation
Sterile, hydrated and ready to use

Removes up to 99.999% salts, dyes, haptens and other small molecules
Samples up to 100 µl are processed in under 5 minutes

Unique conical column for better purification

CentriPure MINI Spin Columns are used for rapid desalting, buffer exchange, or removal of small molecules.

The unique conical column design allows purification of samples up to 100 µl. Salts, dyes, haptens, and dideoxy terminators are completely removed in under 5 minutes. The columns are sterile and ready-to-use.

CentriPure MINI Spin Columns are available pre-swollen in either deionized water, TRIS or PBS. Columns are pre-packed with either **Zetadex-25** (MWCO 5 kD/10 bases) or **Zetadex-50** (MWCO 25 kD/20 bases).

CentriPure MINI Spin Columns are available in kits of 4, 25 or 100 columns.

quick simple efficient

engineered molecular precision

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Unique conical column for greater separation. Sterile, hydrated and ready to use

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Centri Pure MINI:	Desalt Z-50	Desalt Z-25	Desalt Z-25AZ	SEQ Z-50	TRIS Z-50	TRIS Z-25	PBS Z-50	PBS Z-25	BOR Z-25	BOR Z-50	TE Z-50	STE Z-50	SSC Z-50	SSC Z-25
Application:	For rapid desalting in under 5 minutes	For rapid desalting in under 5 minutes Smaller gel pore size	For rapid desalting in under 5 minutes Stabilized with sodium azide Smaller gel pore size	Optimized for nucleic acid sequencing. For removal of up to 99.999% salts, dyes and dideoxy terminators	Pre-hydrated with TRIS buffer. For purification and buffer exchange into TRIS	Pre-hydrated with TRIS buffer. For purification and buffer exchange into TRIS Smaller gel pore size	Pre-hydrated with PBS buffer. For purification and buffer exchange into PBS	Pre-hydrated with PBS buffer. For purification and buffer exchange into PBS Smaller gel pore size	Pre-hydrated with 50 mM Sodium Borate, pH 8. For purification and buffer exchange into Borate Buffer.	Pre-hydrated with 50 mM Sodium Borate, pH 8. For purification and buffer exchange into Borate Buffer.	Pre-hydrated with TE buffer, 10 mM Tris-HCl, 1.0 mM EDTA, pH 8. For purification and buffer exchange into TE Buffer, pH 8.0.	Pre-hydrated with STE buffer, 10 mM Tris-HCl, 1.0 mM EDTA, 0.1 M NaCl, pH 8. For purification and buffer exchange into STE Buffer, pH 8.0.	Pre-hydrated with SSC buffer, 150 mM NaCl, 15 mM sodium citrate, pH 7. For purification and buffer exchange into SSC Buffer, pH 7.0.	Pre-hydrated with SSC buffer, 150 mM NaCl, 15 mM sodium citrate, pH 7. For purification and buffer exchange into SSC Buffer, pH 7.0.
For processing oligonucleotides:	> 20 bases	> 10 bases	> 10 bases	> 20 bases	> 20 bases	> 10 bases	> 20 bases	> 10 bases	> 10 bases	> 20 bases	> 20 bases	> 20 bases	> 20 bases	> 10 bases
For processing proteins:	> 25 kD	> 5 kD	> 5 kD	> 25 kD	> 25 kD	> 5 kD	> 25 kD	> 5 kD	> 5 kD	> 25 kD	> 25 kD	> 25 kD	> 25 kD	> 5 kD
Gel Matrix:	Zetadex-50SF in deionized water	Zetadex-25SF in deionized water	Zetadex-25SF in deionized water and 0.02% sodium azide	Zetadex-50SF in deionized water	Zetadex-50SF in 1 mM TRIS, pH 8	Zetadex-25SF in 1 mM TRIS, pH 8	Zetadex-50SF in standard PBS, pH 7	Zetadex-25SF in standard PBS, pH 7	Zetadex-25SF in 50 mM NaB(OH)4, pH 8	Zetadex-50SF in 50 mM NaB(OH)4, pH 8	Zetadex-50SF in TE buffer, pH 8	Zetadex-50SF in STE buffer, pH 8	Zetadex-50SF in SSC buffer, pH 7	Zetadex-25SF in SSC buffer, pH 7
Gel Bed Volume:	0.5 ml	0.5 ml	0.35 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml	0.5 ml
Optimal Sample Volume:	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl	50 µl
Optimal Centrifuge Conditions:	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.	1000 x g for 2 min.
Removal of Dye: (50 µl 1mM 5/6 carboxyfluore-scein in 0.1M NaHCO¬3)	> 99.9995 %	> 99.99 %	> 99 %	> 99.999 %	> 99.999 %	> 99.95 % (TAMRA dye substituted for fluorescein)	> 99.999 %	> 99.99 %	> 97 %	> 99.9 %	n. a.	> 99.9 %	n. a.	n. a.
Removal of Dye: (100 µl 1mM 5/6 carboxyfluore-scein in 0.1M NaHCO¬3)	> 99.95 %	> 99.5 %	Not recommended to use samples > 50 µl	> 99.95 %	> 99.99 %	> 99.5 % (TAMRA dye substituted for fluorescein)	> 99.99 %	> 99.5 %	> 97 %	> 99.9 %	n. a.	> 99 %	n. a.	n. a.
Removal of Salt: (50 µl 0.8 M NaCl)	> 99.9 % > 99.999 % (with extra wash step *)	> 99.5 %	Not evaluated due to sodium azide	> 99.9 %	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.
Removal of Salt: (100 µl 0.8 M NaCl)	> 99.0 % > 99.5 % (with extra wash step *)	> 99.0 %	Not evaluated due to sodium azide	> 99.0 %	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.	n. a.
Recovery:	Will vary depending on sample and buffer conditions.	Will vary depending on sample and buffer conditions.	> 85 % (50 µl of 1 mg/ml Dextran blue in 0.8 M NaCl)	Will vary depending on sample and buffer conditions.	> 90 % (100 µl 1 mg/ml Dextran blue in water)	> 80 % (100 µl 1 mg/ml Dextran blue in water)	> 85 % (50 µl of 500 µg IgG in PBS/10 % DMSO)	> 80 % (50 µl of 500 µg IgG in PBS/10 % DMSO)	> 80 % (500µg IgG in 50µl PBS)	> 80 % (500µg IgG in 50µl PBS)	> 90 % (1 mg/ml Dextran blue in 100 µl water)	> 90 % (1 mg/ml Dextran blue in 100 µl water)	> 80 % (1 mg/ml Dextran blue in 100 µl water)	> 90 % (1 mg/ml Dextran blue in 100 µl water)
Catalog No.:	CP-0201	CP-0205	CP-0209	CP-0202	CP-0203	CP-0207	CP-0204	CP-0208	CP-0210	CP-0211	CP-0212	CP-0213	CP-0214	CP-0215
Pack Sizes:	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns	4, 25,100 columns

Special or custom resin and buffer filling available on request.
Please contact us at **+49 30 9489 2201** or **tech@empbiotech.com**

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1. Column Preparation

a) Allow the columns at least 15 minutes to warm to room temperature.

b) Tap gently or briefly vortex to resuspend gel and remove air bubbles.

c) Remove bottom cap and then remove top cap.

Figure 1
Orientation mark on column is aligned upward for storage buffer elution.

2. Removal of storage buffer

a) Place the column into a wash tube.

b) Centrifuge at 1000 x g for 2 minutes. Note the column position using the orientation mark (see Fig 1.).

c) Discard wash tube and eluted storage buffer.

d) OPTION: For greater desalting efficiency using Desalt Z-50, add 400 µl distilled water to the column and repeat steps 2a – 2c.

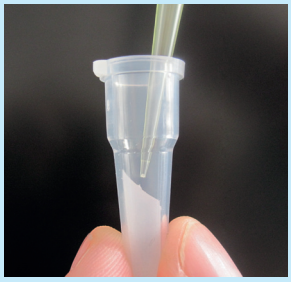

Figure 2
Proper sample application is made directly in to the center of the gel bed.

3. Sample processing

a) Carefully apply sample directly to center of gel bed (see Fig. 2).

b) Place column into a collection tube. Maintain proper column orientation (see Fig 3).

c) Centrifuge at 1000 x g for 2 minutes to elute the purified sample.

Figure 3
Orientation marks are aligned upward for sample elution.

For more information, please contact us at **+49 30 9489 2201**
or **tech@empbiotech.com**

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